

# The Human Immune Response to Dengue Virus Is Dominated by Highly Cross-Reactive Antibodies Endowed with Neutralizing and Enhancing Activity

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## SUMMARY

Antibodies protect against homologous Dengue virus (DENV) infection but can precipitate severe dengue by promoting heterotypic virus entry via Fc $\gamma$  receptors (Fc $\gamma$ R). We immortalized memory B cells from individuals after primary or secondary infection and analyzed anti-DENV monoclonal antibodies (mAbs) thus generated. MAb to envelope (E) protein domain III (DIII) were either serotype specific or cross-reactive and potently neutralized DENV infection. DI/DII- or viral membrane protein prM-reactive mAbs neutralized poorly and showed broad cross-reactivity with the four DENV serotypes. All mAbs enhanced infection at subneutralizing concentrations. Three mAbs targeting distinct epitopes on the four DENV serotypes and engineered to prevent Fc $\gamma$ R binding did not enhance infection and neutralized DENV in vitro and in vivo as postexposure therapy in a mouse model of lethal DENV infection. Our findings reveal an unexpected degree of cross-reactivity in human antibodies against DENV and illustrate the potential for an antibody-based therapy to control severe dengue.

## INTRODUCTION

Dengue virus (DENV) is a mosquito-borne *Flavivirus* responsible for tens of millions of human cases of dengue annually, including 500,000 hospitalizations and 20,000 deaths (Gibbons and Vaughn, 2002), with an economic burden rivaling that of malaria.

A primary infection is believed to provide effective, durable, and possibly life-long protection against reinfection with the same serotype but only short-term protection against other serotypes (Rothman, 2004). Classical epidemiologic studies suggested that immunity to one of the four DENV serotypes can increase disease severity upon subsequent challenge with a different serotype, leading, in some cases, to severe dengue, a disease characterized by plasma leakage and hemorrhagic manifestations (Halstead, 1970). Poorly neutralizing cross-reactive antibodies raised in response to a previous serotype are believed to contribute to pathogenesis of severe dengue by promoting virus entry via Fc $\gamma$  receptors (Fc $\gamma$ R) and infection of myeloid cells (Halstead, 2003), leading to antibody-dependent enhancement (ADE) of infection. The role of antibodies in severe dengue is supported by epidemiological studies showing that infants with waning levels of maternal antibodies (age 6–9 months) are most vulnerable to severe DENV disease (Halstead et al., 2002; Nguyen et al., 2004) and that serum from these infants enhances DENV infection in vitro (Chau et al., 2008; Kliks et al., 1988). The difficulty of balancing immunity to the four serotypes and minimizing incomplete response and the risk of ADE are major hurdles in the development of a tetravalent vaccine against DENV (Whitehead et al., 2007).

The 10.7 Kb RNA genome of DENV encodes three structural proteins, the capsid protein (C), a membrane-associated protein (prM), and an envelope protein (E), and seven nonstructural proteins (NS1, NS2A, NS2B, NS3, NS4A, NS4B, and NS5). The E protein is structurally conserved among flaviviruses and consists of three distinct domains. Domain I (DI) participates in the conformational changes required for viral entry and nucleocapsid escape from the endosomal compartment, domain II (DII) contains the fusion loop, and domain III (DIII) has been suggested to bind cellular receptors (Bhardwaj et al., 2001; Chin et al., 2007; Chu et al., 2005; Rey et al., 1995). Partially mature

**Table 1. DENV-Immune Donors and B Cell Repertoire Analysis**

	Infecting Serotype	Time p.i.	Immortalization Efficiency	Primary Screening	Positive Cultures/Total	DENV- or E-Reactive B Cells (%) <sup>a</sup>
Donor 7	primary DENV-4	200 days	9.1%	ELISA E4 protein	81/2844	1.5%
Donor 13 <sup>b</sup>	primary DENV-2	> 8 years	6.5%	staining of infected C6/36 cells <sup>c</sup>	567/2016	10.7%
				ELISA E2 protein	152/2016	2.9%
Donor 76	primary DENV-3	241 days	14%	staining of infected C6/36 cells <sup>d</sup>	434/960	16%
Donor 12 <sup>e</sup>	secondary DENV-1	510 days	13.8%	staining of infected C6/36 cells <sup>d</sup>	245/1632	5.4%
			12.7%	ELISA E1 protein	313/13824	3.2%
Donor 92	secondary DENV-2	212 days	9%	ELISA DENV lysates	344/2208	8.6%

PBMC were obtained from four donors after a primary or secondary DENV infection and frozen until the day of use. Shown is the infecting serotype and the days postinfection (p.i.) when blood was collected. IgG<sup>+</sup> memory B cells were immortalized with EVB and CpG. Also shown is the efficiency of immortalization and the number and fraction of cultures containing antibodies against DENV identified using different screening methods.

<sup>a</sup> Frequency of DENV- or E-reactive IgG<sup>+</sup> B cells was calculated from the number of positive cultures and the number of input cells per well after correction for the efficiency of B cell immortalization.

<sup>b</sup> Supernatants were tested using two different screening assays.

<sup>c</sup> C6/36 cells were infected with DENV-2, WHO reference strain 516803.

<sup>d</sup> C6/36 cells were infected with DENV isolated from the same donor.

<sup>e</sup> Two independent immortalizations were performed.

virions also express varying levels of prM protein on their surface, which is normally cleaved by a furin-like cellular protease to generate the mature virion (Stadler et al., 1997).

The most potent neutralizing antibodies against DENV, or other flaviviruses such as West Nile Virus (WNV), bind to DIII and have been shown in some cases to be effective as passive prophylaxis or therapy in rodents (Beasley and Barrett, 2002; Goncalvez et al., 2008; Gromowski et al., 2008; Kaufman et al., 1987; Oliphant et al., 2005; Sánchez et al., 2005; Shrestha et al., 2010; Sukupolvi-Petty et al., 2007). DIII-reactive antibodies produced by mice immunized with virus and boosted with recombinant E protein are largely serotype-specific and do not neutralize all of the genotypes within a given serotype (Shrestha et al., 2010). The role of antibodies to DI/DII is less clear, as they tend to be more cross-reactive and less potent in neutralization (Crill and Chang, 2004; Goncalvez et al., 2004; Oliphant et al., 2006). Antibodies to prM generally have poor neutralizing and enhancing activity (Falconar, 1999; Huang et al., 2006), although recent studies suggest that some anti-prM mAbs can augment infectivity of poorly infectious immature virions (Rodenhuis-Zybert et al., 2010). Antibodies against NS1, a secreted nonstructural glycoprotein that is absent from the virion but expressed on the cell surface, can also protect against infection in vivo through FcγR-dependent and -independent mechanisms (Chung et al., 2006) or can possibly contribute to pathogenesis (Falconar, 2007; Lin et al., 2008).

Our current knowledge of the human antibody response to DENV is mostly based on serological studies. In this study, we used an improved method of memory B cell immortalization (Traggiai et al., 2004) combined with a broad screening approach to isolate a large panel of DENV-reactive mAbs from human donors. MAb that bound to DIII of the E protein potently neutralized DENV infection, although they also showed enhancing activity at lower concentrations. In contrast, mAbs that bound to DI/DII of E protein or prM neutralized poorly yet potently

enhanced infection of FcγR-bearing cells. Surprisingly, some of the human DIII-specific mAbs and all DI/DII-specific mAbs isolated from secondary infection showed a broad pattern of cross-reactivity with the four DENV serotypes. Based on an improved understanding of the functional humoral response against DENV, we produced a cocktail of three variant recombinant mAbs that neutralized all four DENV serotypes without causing immune enhancement in vitro and in vivo.

## RESULTS

### High Frequency of DENV-Specific Memory B Cells in Immune Donors

Blood samples were collected from donors at different time points after diagnosis of either primary (donor 7, donor 13, and donor 76) or secondary (donor 12 and donor 92) DENV infection. IgG<sup>+</sup> memory B cells isolated from frozen PBMC were immortalized with EBV and CpG in multiple replicate wells as previously described (Traggiai et al., 2004). Culture supernatants were collected after 2 weeks and analyzed by staining of fixed and permeabilized C6/36 mosquito cells infected with DENV or by ELISA using recombinant E protein from DENV-1, DENV-2, and DENV-4 or lysates of DENV-infected cells (Table 1). The frequency of IgG<sup>+</sup> memory B cells reactive against DENV-infected cells, calculated from the number of positive cultures and the number of input cells after corrections for the efficiency of B cell immortalization, was high in all donors, ranging from 5.4% to 16% of IgG<sup>+</sup> memory B cells. In the three donors that were screened using E protein, the frequency of E-reactive IgG<sup>+</sup> memory B cells ranged from 1.5% to 3.2%. These findings indicate that DENV-immune donors maintain a very large pool of memory B cells reactive against DENV proteins, even several years after infection.

Because the E protein, and in particular DIII, is the main target of neutralizing anti-WNV antibodies in mice (Oliphant et al.,

2007), we performed a large screen to gain insights into the domain specificity and cross-reactivity of E-specific antibodies isolated from donor 13 (primary DENV-2) and donor 12 (secondary DENV-1). Eighteen out of 152 antibodies from donor 13 bound to DIII of DENV-2. Of these, 13 were serotype specific, 1 cross-reacted with DENV-2, -3, and -4, and 4 cross-reacted with all four DENV serotypes. In the case of donor 12, a large fraction of E-reactive antibodies bound to DIII (138 out of 313). Of these, 44 were specific for DIII of DENV-1, whereas the remaining 94 were cross-reactive with two, three, or all four DENV serotypes (16, 15, and 63, respectively). Taken together, these findings indicate that the human antibody response to DIII of DENV E protein elicited by natural infection comprises serotype-specific as well as broadly cross-reactive antibodies.

### Isolation of Human Monoclonal Antibodies to DENV

In order to better characterize the human antibody response to DENV, we isolated B cell clones from several independent cultures and characterized 70 DENV-reactive monoclonal antibodies (mAbs) (68 IgG1, either  $\kappa$  or  $\lambda$ , 1 IgG3,  $\lambda$ , and 1 IgG4,  $\lambda$ ). All DENV-reactive mAbs bound to one or more DENV serotypes, as shown by staining of Vero cells infected with DENV-1, DENV-2, DENV-3, or DENV-4 (see examples in Figure 1A). Specificity and cross-reactivity were assessed using binding assays (ELISA with recombinant E, NS1, and NS3 proteins, staining of yeast displaying DI/DII or DIII, or western blot of infected cell lysates) (see examples in Figures 1B–1D). Mabs were also characterized using functional assays (neutralization or enhancement of infection by DENV vaccine strains using flow cytometry assays with Vero and K562 cells, respectively; Kraus et al., 2007; Littau et al., 1990) (see examples in Figures 1E and 1F).

### Serotype-Specific and Cross-Reactive mAbs to DIII of E Protein

Of the 70 DENV-reactive mAbs isolated, 13 mapped to DIII of E protein, and of these, 5 were serotype specific, whereas 8 were cross-reactive with two, three, or even all four DENV serotypes (Table 2). Of the five serotype-specific mAbs, three (DV3.7, DV25.5, and DV470.12) potentially neutralized infection of Vero cells by either DENV-1 or DENV-2 vaccine strains, with EC<sub>50</sub> values in the low ng/ml range, whereas two mAbs (DV1.6 and DV14.5) showed decreased potency against DENV-2, suggesting that they may represent low-affinity antibodies or may target private epitopes within DENV-2. Of the eight cross-reactive mAbs, two (DV63.1 and DV55.1) potentially neutralized DENV-1 and DENV-3, whereas mAb DV87.1 potentially neutralized DENV-1, DENV-2, and DENV-3. Strikingly, the cross-reactive mAbs DV63.1 and DV87.1 neutralized infection in the same range of concentrations as observed for serotype-specific mAbs (Table 2). Finally, four mAbs bound all serotypes by ELISA, and of these, two neutralized, albeit with low potency, all serotypes (DV21.5 and DV257.13), whereas the remaining mAbs showed a more restricted pattern of neutralization.

Consistent with what is known about the stoichiometric relationship between neutralization and enhancement (Pierson et al., 2007), DIII-specific mAbs also enhanced infection of K562 cells over a given range of concentrations with the

same serotype specificity as shown in the neutralization assay (Table 2).

These results indicate that it is possible to isolate DIII-reactive human mAbs that potentially neutralize infection of two or more DENV serotypes.

### DI/DII-Specific mAbs from Secondary DENV Infection Cross-React with All Four DENV Serotypes

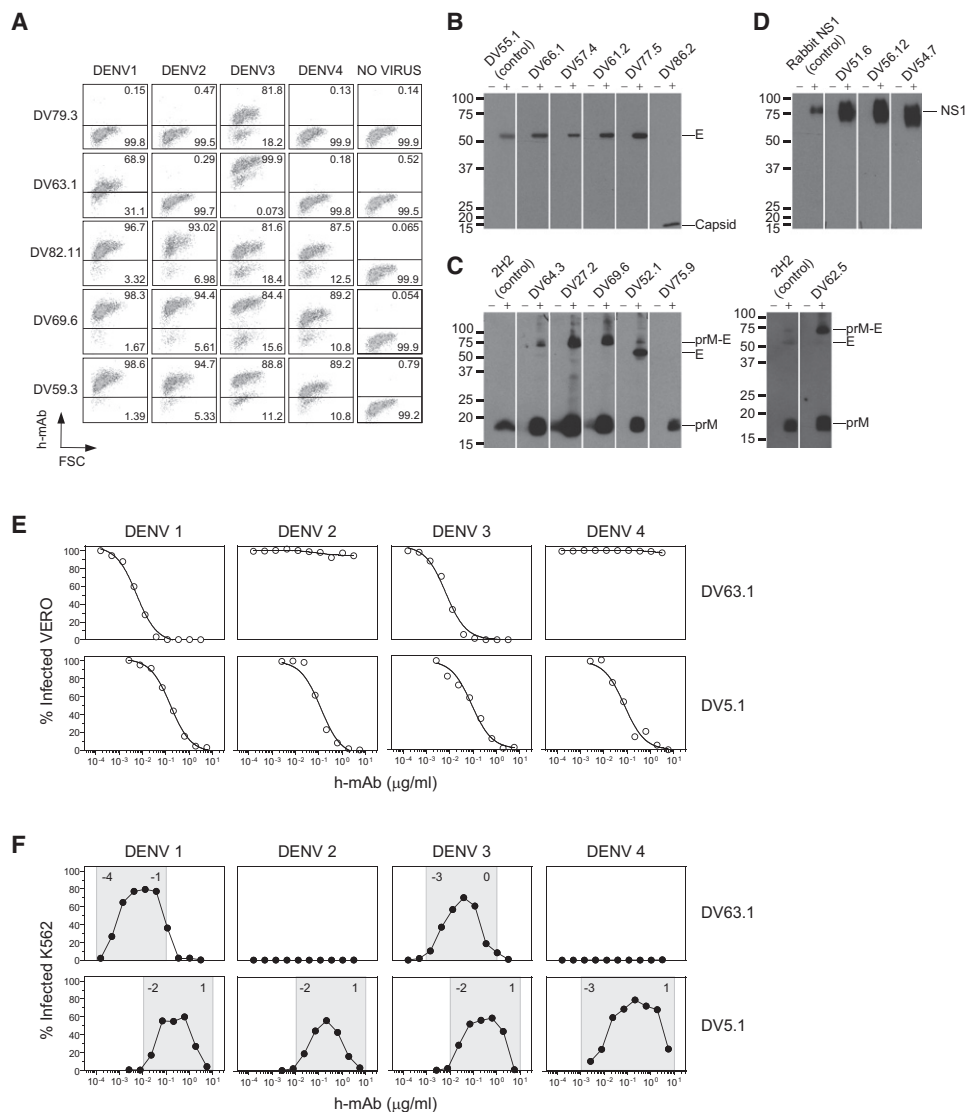
Of the 70 DENV-reactive mAbs, 34 mapped to DI/DII of E protein (Table 3). Of these, 8 were serotype-specific, 1 cross-reacted with DENV-1 and DENV-2, and 25 cross-reacted with all four DENV serotypes. When compared to DIII-specific mAbs, the DI/DII-specific mAbs showed in general a 10- to 50-fold lower potency of neutralization and enhanced infection over a wide range of concentrations (Table 3). Of interest, whereas DI/DII-reactive mAbs isolated from primary infections (donors 7, 13, and 76) were mostly serotype specific, all DI/DII-reactive mAbs isolated from secondary infection (donors 12 and 92) neutralized and enhanced infection by all four DENV serotypes. These findings suggest that broadly cross-reactive DI/DII-specific memory B cells may be selected during the course of secondary DENV infection.

Some of the broadly reactive DI/DII-specific mAbs were analyzed for reactivity against West Nile virus (WNV). Seven mAbs stained yeast cells expressing DI/DII of WNV E protein (Table S1 available online). Using site-directed mutants in DII that had been previously generated (Crill and Chang, 2004; Oliphant et al., 2006), we localized binding of three mAbs to the fusion loop peptide at the tip of DII (W101R, G106E) and of a fourth mAb to the W233F residue on the dimmer interface of DII. The binding of the other three cross-reactive mAbs was not affected by these mutations, indicating that they map to DI/DII but outside of the fusion loop region.

Finally, 4 mAbs (DV66.1, DV57.4, DV61.2, and DV77.5), which were selected for binding to DENV-infected cells, did not bind to recombinant E proteins in ELISA but bound to E protein in western blot (Figure 1B). DV66.1 and DV57.4 neutralized one and three DENV serotypes, respectively, whereas DV61.2 and DV77.5 did not neutralize DENV infection at the highest concentration tested. However, all 4 mAbs showed ADE activity (Table 3).

### Human Monoclonal Antibodies to prM and to Nonstructural Proteins

We next characterized 19 mAbs that stained DENV-infected cells but did not recognize E protein. Six mAbs reacted with a band in western blot corresponding to prM, as indicated by the mouse anti-prM mAb 2H2 used as control (Falconar, 1999) (Figure 1C). As independent verification, we also performed cross-competition experiments and found that the anti-prM mAbs DV64.3, DV27.2, and DV69.6 inhibited binding of 2H2, suggesting that they may recognize the same or overlapping epitopes, whereas DV52.1, DV75.9, and DV62.5 did not compete with 2H2 and may recognize distinct epitopes (data not shown). Surprisingly, five of these mAbs stained a band corresponding to E-prM protein, and three of them (DV64.3, DV52.1, and DV62.5) also recognized a band corresponding to the E protein. The reactivity of mAbs with both prM and E suggests a quaternary epitope with shared sites on the heterodimeric prM-E protein;



**Figure 1. Specificity and Function of Human mAbs against DENV**

(A) Vero cells, uninfected or infected by DENV-1, -2, -3, and -4 vaccine strains, were fixed, permeabilized, and stained with human mAbs, followed by fluorescently labeled anti-human IgG polyclonal antibodies. Representative dot plots show staining by mAbs isolated from donor 76 reactive against DENV-3 alone (mAb DV79.3), DENV-1 and DENV-3 (mAb DV63.1), or cross-reactive against the four DENV serotypes (mAbs DV82.11, DV69.6, DV59.3). Numbers in quadrants indicate the percentage of positive cells. The data shown are for five mAbs and are representative of data obtained with 70 DENV-reactive mAbs and of at least two independent experiments.

(B–D) Western blot analysis of mAbs that were not reactive by ELISA. Concentrated supernatants of uninfected control (–) or DENV-3-infected Vero cells (+) were separated under nonreducing conditions. (B) E- and capsid-specific mAbs. E protein was identified by DV55.1, a mAb specific for E protein in ELISA. Capsids migrate with apparent molecular weights of 16 kD. (C) prM-specific mAbs. prM protein was identified by the mouse anti-prM mAb 2H2. (D) NS1-specific mAbs. NS1 protein was identified by a rabbit anti-NS1 serum. The data are representative of at least two independent experiments.

(E) Examples of viral neutralization assay in which serial dilutions of mAbs (DV63.1 and DV5.1) were incubated with DENV-1–4 before addition to Vero cells. Shown is the percentage of infected cells after 3 days as a function of increasing mAb concentration. Nonlinear regression analysis was used to calculate the  $\text{EC}_{50}$  values indicated in the panel.

(F) Infection enhancement assay in which serial mAb dilutions were incubated with DENV-1–4 before addition to K562 cells. Shown is the percentage of infected K562 cells. The range of mAb concentrations where infection enhancement was observed is indicated in gray. Numbers in the gray areas represent logarithm power. The data shown are from one representative experiment out of three independent experiments performed. Data obtained with the panel of DENV-reactive mAbs are shown in Tables 2, 3, and 4.

a similar finding has been reported for mouse antibodies (Falconar, 1999; Gould et al., 1985; Puttikhunt et al., 2008). With the single exception of mAb DV52.1, all of the prM-specific mAbs were broadly cross-reactive, as they bound to cells

infected with all four DENV serotypes (Table 4). All prM-reactive mAbs enhanced DENV infection over a broad range of concentrations, whereas only three neutralized infection with low potency (Table 4). Thus, consistent with a recent study



**Table 2. Characterization of DIII-E-Reactive Human mAbs**

mAbs	Donor	Isotype	Specificity	ELISA E Protein				Yeasts DV2				Neutralization (EC <sub>50</sub> µg/ml)				Range of Enhancement (log <sub>10</sub> µg/ml)			
				DV 1	DV 2	DV 3	DV 4	DIII	DI/DII	DIII	DIII	DV 1	DV 2	DV 3	DV 4	DV 1	DV 2	DV 3	DV 4
DV 1.6	Don 13	γ1, κ	E, DIII	+	+	+	+	+	n.d.	+	+	+	3.5	—	—	—	—	—	—
DV 3.7	Don 13	γ1, λ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.003	—	—	—	—3, 1	—	—
DV 14.5	Don 13	γ1, λ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.043	—	—	—	—1, 1	—	—
DV 25.5	Don 13	γ1, κ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.005	—	—	—	—3, 1	—	—
DV 470.12	Don 12	γ1, λ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.002	—	—	—3, 1	—	—	—
DV 63.1	Don 76	γ1, κ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.006	—	0.006	—4, —1	—	—3, 0	—
DV 55.1	Don 76	γ1, λ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.013	0.577	0.014	—3, 0	—1, 1	—3, 1	—
DV 87.1	Don 12	γ1, κ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.004	0.004	0.008	—3, 1	—2, 1	—2, 0	—
DV 291.7	Don 12	γ1, κ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.147	0.056	0.820	—4, 0	—2, 0	—2, 0	—
DV 10.16	Don 13	γ1, λ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.083	—	0.411	0, 3	—2, 1	—1, 1	—2, 1
DV 21.5	Don 13	γ1, λ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.410	1.011	0.726	—1, 2	—1, 3	—1, 3	0, 3
DV 257.13	Don 12	γ1, κ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.107	0.154	0.193	n.d.	n.d.	n.d.	n.d.
DV 415.8	Don 12	γ1, κ	E, DIII	+	+	+	+	+	n.d.	+	+	—	0.134	0.163	0.127	n.d.	n.d.	n.d.	n.d.

The table reports the phenotype of DIII-E-specific mAbs. The target specificity of the mAbs was determined by ELISA. The donor from which the mAbs were isolated and the mAb isotype and light-chain usage are indicated. Viral neutralization was performed using Vero cells and DENV-1-4 vaccine strains. Shown are EC<sub>50</sub> values. Enhancement of infectivity was performed using K562 cells. Shown is the range of mAb concentrations for which infection enhancement was observed (see example in Figure 1F). The data shown are representative of two independent experiments. n.d., not done. (+) OD values > 0.5. (–) OD values < 0.5 or no neutralization at the highest concentration tested (5 or 10 µg/ml).

(Dejnirattisai et al., 2010), the prM-specific mAbs were highly cross-reactive among the DENV serotypes and potentially promoted ADE.

Eleven mAbs were mapped to NS1 or NS3 protein, as determined by ELISA with recombinant proteins and western blot (Figure 1D and data not shown). Five NS1-specific mAbs cross-reacted with all DENV serotypes, whereas the remaining two mAbs reacted with DENV-1 and DENV-3, and the four NS3-specific mAbs were all DENV-3-specific (Table 4). Finally, only one mAb (DV86.2) recognized the capsid protein (Table 4 and Figure 1B). Another mAb (DV34.4) was not assigned to a specific protein, although it showed high neutralizing activity against DENV-2 and, to a lower extent, DENV-4 (Table 4). This mAb likely recognizes a conformationally sensitive epitope on E (e.g., hinge or interface between domains of oligomers), as was observed recently with two human mAbs against WNV (Vogt et al., 2009).

### A Broadly Neutralizing Antibody Cocktail Lacking Enhancing Activity

Given the availability of human mAbs that potentially neutralized DENV infection, we set out to develop a candidate antibody-based therapeutic for dengue consisting of a “cocktail” of neutralizing mAbs. Ideally, this cocktail should: (1) neutralize all DENV serotypes, (2) target at least two nonoverlapping epitopes on each virus in order to minimize selection of escape mutants, and (3) fail to enhance infection. We therefore selected two mAbs with potent and complementary neutralizing activity: DV87.1 specific for DIII of DENV-1, -2, and -3 and DV22.3 specific for DI/DII of DENV-4. As a complement, we selected a DI/DII-specific mAb (DV82.11) that neutralized with comparable efficiency all four DENV serotypes (Figure 2A). Cross-competition experiments were consistent with the notion that the target epitopes recognized by the three mAbs did not overlap (Figure S1). Thus, a cocktail of DV87.1, DV22.3, and DV82.11 is expected to neutralize all DENV serotypes by targeting two distinct sites on each virus.

Because the selected neutralizing mAbs also enhanced infection (Figure 2A), we generated variants of DV87.1, DV22.3, and DV82.11 in which leucine residues at positions 1.3 and 1.2 of CH2 domain (according to the IMGT unique numbering for C domain) were substituted with alanine residues. This modification, also known as “LALA” mutation, abolishes antibody binding to FcγRI, FcγRII, and FcγRIIIa (Hessell et al., 2007). The variant and unmodified recombinant mAbs were expressed in HEK293T cells, and the purified mAbs were compared for their capacity to neutralize and enhance infection by the four DENV serotypes. As shown in Figure 2B, LALA variants retained the same neutralizing activity as unmodified mAbs but were completely devoid of enhancing activity. In addition, the LALA cocktail, even at very low concentrations (DV82.11 = 2 µg/ml; DV87.1 = DV22.3 = 0.2 µg/ml), blocked ADE induced by enhancing prM or DI/DII mAbs or immune serum (Figure 2C).

To determine whether the combination of LALA mAbs might achieve increased neutralization, we used a variable ratio approach (Zwick et al., 2001). The concentration of one mAb was varied in the presence of a fixed amount of a second mAb (added at a weakly neutralizing concentration), and the titration curves of the first mAb alone or in combination were compared.

**Table 3. Characterization of DI/DII E-Reactive Human mAbs**

mAbs	Donor	Isotype	Specificity	ELISA E Protein						Yeasts DV2		Neutralization (EC <sub>50</sub> µg/ml)				Range of Enhancement (log10 µg/ml)			
				DV 1	DV 2	DV 3	DV 4	DIII	DV 3/4	DI/DII	DIII	DV 1	DV 2	DV 3	DV 4	DV 1	DV 2	DV 3	DV 4
DV 13.8	Don 13	γ1, λ	E, DI/DII	—	+	—	—	—		n.d.	—	—	0.241	—	—	—	–2, 1	—	—
DV 16.8	Don 7	γ1, κ	E, DI/DII	—	—	—	+	—		n.d.	—	—	—	—	0.039	—	—	—	–2, 0
DV 18.21	Don 13	γ1, κ	E, DI/DII	—	+	—	—	—		n.d.	—	—	0.003	—	—	—	–3, 0	—	—
DV 22.3	Don 7	γ1, λ	E, DI/DII	—	—	—	+	—		n.d.	—	—	—	—	0.006	—	—	—	–2, 1
DV 35.3	Don 13	γ1, κ	E, DI/DII	—	+	—	—	—		n.d.	—	—	0.918	—	—	—	–1, 1	—	—
DV 74.4	Don 76	γ1, λ	E, DI/DII	—	—	+	—	—		n.d.	—	—	—	0.020	—	—	—	–2, 0	—
DV 79.3	Don 76	γ1, κ	E, DI/DII	—	—	+	—	—		n.d.	—	—	—	0.023	—	—	—	–2, 0	—
DV 76.5	Don 76	γ1, κ	E, DI/DII	—	—	+	—	—		n.d.	—	—	—	—	—	–1, 2	–2, 1	–2, 1	–2, 1
DV 90.3	Don 76	γ1, λ	E, DI/DII	+	—	+	—	—		n.d.	—	0.342	—	0.454	—	–2, 1	—	–1, 1	—
DV 1.1	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.209	0.066	0.262	0.183	–2, 1	–2, 1	–2, 1	–2, 1
DV 4.4	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.248	0.214	0.167	0.760	–2, 1	–2, 1	–1, 1	–2, 1
DV 5.1	Don 92	γ1, κ	E, DI/DII	+	+	+	+	—		+	—	0.156	0.103	0.092	0.069	–2, 1	–2, 1	–2, 1	–3, 1
DV 6.1	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.382	0.296	0.351	0.170	–1, 1	–2, 1	–1, 1	–2, 1
DV 7.5	Don 92	γ1, κ	E, DI/DII	+	+	+	+	—		+	—	0.084	0.075	0.130	0.031	–2, 1	–2, 1	–2, 1	–2, 1
DV 8.1	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.103	0.022	0.031	0.024	–3, 2	–2, 1	–3, 1	–3, 0
DV 13.4	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.063	0.023	0.016	0.045	–2, 1	–3, 0	–3, 0	–3, 0
DV 14.5	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.053	0.049	0.060	0.040	–2, 0	–2, 0	–2, 0	–3, 0
DV 15.7	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.123	0.055	0.173	0.070	–2, 1	–3, 1	–2, 1	–3, 1
DV 16.5	Don 92	γ1, κ	E, DI/DII	+	+	+	+	—		+	—	0.248	0.089	0.219	0.101	–2, 1	–2, 1	–2, 1	–2, 1
DV 17.6	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.218	0.103	0.256	0.087	–2, 1	–2, 1	–2, 1	–2, 2
DV 18.4	Don 92	γ1, κ	E, DI/DII	+	+	+	+	—		+	—	0.670	0.241	0.799	0.699	–2, 1	–2, 1	–2, 1	–2, 1
DV 19.3	Don 92	γ1, κ	E, DI/DII	+	+	+	+	—		+	—	0.631	0.189	0.368	0.114	–2, 1	–2, 1	–2, 1	–2, 1
DV 20.1	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.244	0.128	0.777	0.226	–2, 1	–2, 1	–1, 1	–2, 1
DV 21.1	Don 92	γ1, κ	E, DI/DII	+	+	+	+	—		+	—	0.161	0.055	0.069	0.101	–2, 1	–2, 1	–3, 1	–3, 1
DV 23.13	Don 13	γ1, λ	E, DI/DII	+	+	+	+	—		n.d.	—	n.d.	0.013	n.d.	n.d.	n.d.	–3, 1	n.d.	n.d.
DV 28.1	Don 12	γ1, λ	E, DI/DII	+	+	+	+	—		n.d.	—	0.093	0.027	0.084	0.027	–3, 1	–3, 1	–3, 1	–3, 1
DV 28.8	Don 92	γ1, κ	E, DI/DII	+	+	+	+	—		+	—	0.106	0.048	0.233	0.092	–2, 0	–2, 0	–2, 0	–2, 1
DV 38.1	Don 92	γ1, λ	E, DI/DII	+	+	+	+	—		+	—	0.111	0.036	0.239	0.106	–2, 1	–3, 1	–2, 1	–3, 1
DV 78.6	Don 76	γ1, κ	E, DI/DII	+	+	+	+	—		n.d.	—	0.591	0.251	0.809	0.367	–2, 1	–2, 1	–2, 1	–2, 1
DV 82.11	Don 76	γ1, λ	E, DI/DII	+	+	+	+	—		n.d.	—	0.043	0.024	0.090	0.117	–2, 1	–2, 1	–2, 1	–2, 1
DV 143.6	Don 12	γ1, κ	E, DI/DII	+	+	+	+	—		n.d.	—	0.253	0.083	0.255	0.070	–2, 1	–2, 1	–2, 1	n.d.
DV 163.3	Don 12	γ1, κ	E, DI/DII	+	+	+	+	—		n.d.	—	0.170	0.100	0.187	0.542	–2, 1	–2, 0	–1, 0	n.d.
DV 378.12	Don 12	γ1, κ	E, DI/DII	+	+	+	+	—		n.d.	—	0.149	0.072	0.251	0.498	–2, 1	–2, 1	–3, 1	n.d.
DV 291.3	Don 12	γ1, λ	E, DI/DII	+	+	+	+	—		n.d.	—	0.120	0.080	0.199	0.202	–2, 0	–2, 1	–2, 1	n.d.
DV 66.1	Don 76	γ1, κ	E	—	—	—	—	—		n.d.	—	—	—	1.950	—	—	—	–2, 1	—

**Table 3. Continued**

mAbs	Donor	Isotype	Specificity	ELISA E Protein				Yeasts DV2		Neutralization (EC <sub>50</sub> µg/ml)				Range of Enhancement (log10 µg/ml)			
				DV 1	DV 2	DV 3	DV 4	DIII DV 3/4	DII/DII	DV 1	DV 2	DV 3	DV 4	DV 1	DV 2	DV 3	DV 4
DV 57.4	Don 76	γ1, κ	E	—	—	—	—	—	n.d.	—	0.200	0.167	0.193	—	—3, 0	—3, 0	—2, 0
DV 61.2	Don 76	γ1, κ	E	—	—	—	—	—	n.d.	—	—	—	—	0, 2	—2, 1	—2, 1	—2, 1
DV 77.5	Don 76	γ1, κ	E	—	—	—	—	—	n.d.	—	—	—	—	—1, 2	—2, 0	—2, 1	—2, 1

The table reports the phenotype of DII/DII- and E-specific mAbs, the donor from which the mAbs were isolated, and the mAb isotype and light-chain usage as determined by specific ELISA. The target specificity of the mAbs was determined by ELISA performed with recombinant E proteins (from DENV-1–4 serotypes) or with DIII of DENV-3 or DENV-4 (DV3/4) and by staining of yeast expressing DII/DIII of DENV-2 E protein. Four mAbs did not recognize E proteins by ELISA but bound E protein in western blot. Shown are EC<sub>50</sub> values and the range of mAb concentrations for which infection enhancement was observed (as in Table 2). The data shown are representative of two independent experiments. n.d., not done. (+) OD values > 0.5. (–) OD values < 0.5 or no neutralization at the highest concentration tested (5 or 10 µg/ml). See also Table S1.

The EC<sub>90</sub> values shown in Table S2 indicate a 2- to 4-fold increase in neutralization potency, consistent with a modest additive effect.

Taken together, the above results suggest that a cocktail of three LALA variant mAbs targeting different epitopes on each virus can efficiently neutralize DENV infection while avoiding and actually preventing enhancement of DENV infection in vitro.

### LALA Variant mAbs Do Not Enhance DENV Infection and Show Therapeutic Efficacy In Vivo

To assess the ability of LALA variants to neutralize DENV in vivo, we utilized a recently described mouse model of DENV ADE in which sublethal infection is enhanced by passive transfer of anti-DENV antibodies, leading to lethal disease characterized by plasma leakage, elevated serum cytokines, and thrombocytopenia (Balsitis et al., 2010; Zellweger et al., 2010), all features associated with severe dengue in humans. Unmodified or LALA variant mAbs were transferred into AG129 mice 24 hr prior to a sublethal infection with DENV-2 strain D2S10. Whereas mice receiving 5 µg of the mouse mAb 4G2 (pan-flavivirus, E DII/DIII specific) developed an enhanced, lethal D2S10 infection, mice receiving 1 or 5 µg of LALA mAb variants did not succumb to infection (Figure 3A; 1 or 5 µg DV87.1 LALA,  $p < 0.0253$ ; 1 or 5 µg DV82.11 LALA,  $p < 0.0455$  as compared to 5 µg 4G2) or show signs of illness. In contrast, with one exception, all of the mice pretreated with 1 or 5 µg unmodified mAbs DV87.1 and DV82.11 showed enhanced lethal DENV infection, whereas five of six mice receiving PBS alone survived infection (Figure 3A). The ability of the unmodified mAbs to enhance DENV in mice was not affected by differences in the interaction between the human mAbs and the murine Fcγ. Indeed, chimeric mAbs in which the human constant region (hinge, CH2 and CH3 domains) was replaced with the homologous murine Igγ2a constant region enhanced DENV infection comparably in vitro and in vivo (data not shown).

To determine whether survival associated with transfer of the DV87.1 and DV82.11 LALA variants was associated with reduced viral load, we measured the viral burden in serum and tissues 3.5 days following D2S10 antibody-enhanced infection. Viremia and tissue viral load measured in liver, small intestine, and lymph node were significantly decreased in mice receiving 5 µg of either LALA variant as compared to mice receiving 5 µg of the parent unmodified mAbs (Figure S2,  $p < 0.0495$  for comparisons between mAb variants in the liver, sera, and lymph; in the small intestine,  $p < 0.0369$  for comparison between DV87.1 mAb variants and  $p < 0.0463$  between DV82.11 mAb variants).

To explore whether the LALA variants could serve as a possible therapy following DENV infection, we administered 50 µg of the LALA variants or unmodified mAbs to mice 24 hr after infection with DENV-2 D2S10 under enhancing conditions (24 hr after transfer of heterotypic anti-DENV-1 serum) (Balsitis et al., 2010). Mice receiving either the DV87.1 or DV82.11 LALA variant survived the normally lethal infection, whereas mice receiving the unmodified parent mAbs succumbed to infection, as did mice receiving a nonbinding isotype control (DV22.3 LALA mAb that only recognizes DENV-4) or PBS (Figure 3B, DV87.1 LALA,  $p < 0.0015$ ; DV82.11 LALA,  $p < 0.011$  compared to DV22.3 LALA). In summary, engineering of the LALA mutation

**Table 4. Characterization of Non-E-Reactive mAbs**

mAbs	Donor	Isotype	Specificity	Staining of Vero-Infected Cells				Neutralization (EC <sub>50</sub> µg/ml)				Range of Enhancement (log <sub>10</sub> µg/ml)			
				DV 1	DV 2	DV 3	DV 4	DV 1	DV 2	DV 3	DV 4	DV 1	DV 2	DV 3	DV 4
DV 64.3	Don 76	γ1, κ	prM	+	+	+	+	0.327	0.402	0.067	0.128	−4, 1	−4, 1	−4, 1	−3, 1
DV 27.2	Don 92	γ1, κ	prM	+	+	+	+	0.391	0.345	0.110	0.076	−3, 1	−3, 1	−3, 1	−3, 1
DV 69.6	Don 76	γ1, λ	prM	+	+	+	+	0.912	1.559	0.129	0.070	−4, 1	−3, 1	−4, 1	−4, 2
DV 52.1	Don 76	γ1, κ	prM	+	—	+	—	—	—	—	—	—	—	−3, 1	—
DV 75.9	Don 76	γ1, κ	prM	+	+	+	+	—	—	—	—	−2, 0	−1, 2	−3, 0	−2, 0
DV 62.5	Don 76	γ1, λ	prM	+	+	+	+	—	—	—	—	−2, 1	−2, 1	−2, 1	−2, 1
DV 51.6	Don 76	γ1, λ	NS1	+	—	+	—	—	—	—	—	—	—	—	—
DV 53.4	Don 76	γ1, λ	NS1	+	—	+	—	—	—	—	—	—	—	—	—
DV 54.7	Don 76	γ1, κ	NS1	+	+	+	+	—	—	—	—	—	—	—	—
DV 56.12	Don 76	γ1, λ	NS1	+	+	+	+	—	—	—	—	—	—	—	—
DV 59.3	Don 76	γ4, λ	NS1	+	+	+	+	—	—	—	—	—	—	—	—
DV 60.3	Don 76	γ1, κ	NS1	+	+	+	+	—	—	—	—	—	—	—	—
DV 70.1	Don 76	γ1, κ	NS1	+	+	+	+	—	—	—	—	—	—	—	—
DV 65.5	Don 76	γ1, κ	NS3	—	—	+	—	—	—	—	—	—	—	—	—
DV 67.9	Don 76	γ1, λ	NS3	—	—	+	—	—	—	—	—	—	—	—	—
DV 68.2	Don 76	γ3, λ	NS3	—	—	+	—	—	—	—	—	—	—	—	—
DV 71.1	Don 76	γ1, λ	NS3	—	—	+	—	—	—	—	—	—	—	—	—
DV 86.2	Don 76	γ1, λ	capsid	+	+	+	+	—	—	—	—	—	—	—	—
DV 34.4	Don 92	γ1, κ	n.a.	—	+	—	+	—	0.010	—	0.431	—	−3, 0	—	−1, 2

The table reports the phenotype of non-E-reactive mAbs, the donor from which the mAbs were isolated, and the mAb isotype and light-chain usage as determined by specific ELISA. Specificity was determined by staining of Vero cells infected with DENV and by western blot assay or ELISA. The specificity of mAb DV34.4 was not assigned (n.a.). The mAb isotype and light-chain usage were determined as described in Table 2. Viral neutralization and enhancement assays were performed on Vero and K562 cells, respectively, using DENV-1-4 vaccine strains. Shown are EC<sub>50</sub> values of neutralization and range of mAb concentrations of infection enhancement. The data shown are representative of two independent experiments.

on strongly neutralizing human mAbs abrogates the capacity for ADE and confers a protective phenotype as a postexposure therapy in mice.

## DISCUSSION

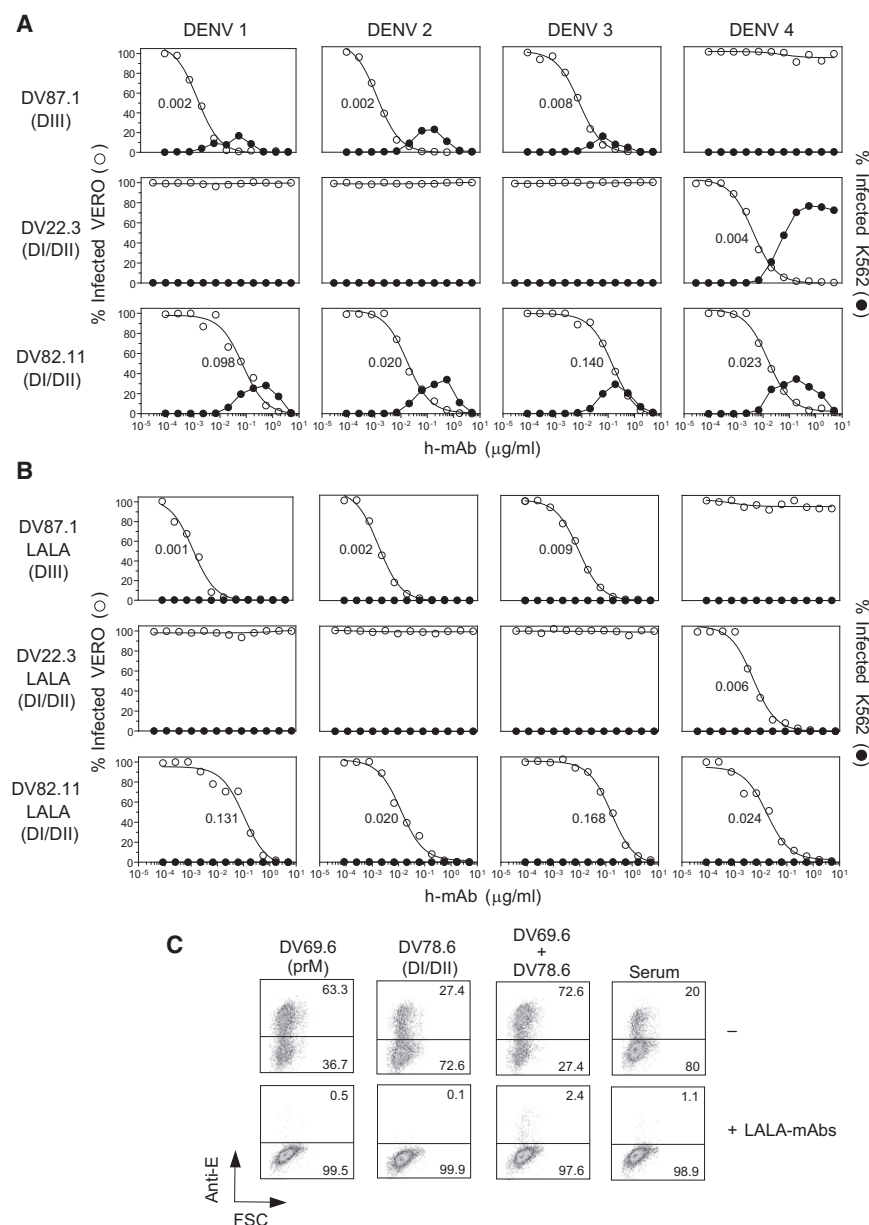
In this study, we analyzed the memory B cell repertoires of individuals recovered from primary or secondary DENV infection and characterized a large panel of mAbs specific for structural and nonstructural proteins of DENV. The main findings are that immune donors have a large repertoire of DENV-reactive memory B cells, even years after infection, and that human E-specific as well as prM-specific mAbs isolated from these donors show extensive cross-reactivity among the four DENV serotypes.

Previous studies using mouse mAbs established that DIII is the primary target of the most potent neutralizing antibodies against flaviviruses, including WNV and DENV (Beasley and Barrett, 2002; Gromowski and Barrett, 2007; Oliphant et al., 2005, 2007; Sánchez et al., 2005; Sukupolvi-Petty et al., 2007). Our findings in humans agree with these observations, as the isolated DIII-specific mAbs were indeed the most potent in neutralizing DENV infection of Vero cells, with EC<sub>50</sub> values in the low ng/ml range. However, whereas mAbs produced by mice boosted with recombinant E protein could only neutralize a few genotypes within a given serotype (Shrestha et al., 2010), the human

mAbs to DIII showed considerable breadth, with some mAbs being able to neutralize three or even all four DENV serotypes. DI/DII-specific mAbs were less potent in viral neutralization and showed an even higher degree of cross-reactivity. Strikingly, analysis performed on a large number of culture supernatants showed that the fraction of DIII and DI/DII cross-reactive antibodies was considerably higher in secondary infections. For instance 44.1% of supernatants containing DIII-specific antibodies from donor 12 (secondary DENV-1) cross-reacted with two, three, or four DENV serotypes, whereas 100% of DI/DII-specific mAbs from donor 92 (secondary DENV-2) cross-reacted with comparable EC<sub>50</sub> values with all four DENV serotypes.

Within the E-specific response, the relative proportion of DI/DII versus DIII antibodies showed considerable variability in different donors. However, there was an overall prevalence of DI/DII antibodies, which is consistent with serological studies with WNV and DENV (Crill et al., 2009; Lai et al., 2008; Oliphant et al., 2007; Wahala et al., 2009). Moreover, two recent studies described recombinant anti-WNV antibodies that were generated after phage display screening of scFv (V<sub>H</sub>-V<sub>L</sub>) molecules (Gould et al., 2005; Throsby et al., 2006), both of which observed immunodominance of DI/DII mAbs against WNV E protein. For unclear reasons, it appears that DIII-specific antibodies are less abundant in the human repertoire, although they still may contribute significantly to protection (Crill et al., 2009).





**Figure 2. Three mAbs Neutralize All Four DENV Serotypes by Binding Two Distinct Epitopes on Each Virus**

Three mAbs (all IgG1) were selected for their specificity and neutralizing activity: DV87.1 binds DIII of DENV-1, -2, and -3; DV22.3 binds DI/DII of DENV-4, and DV82.11 cross-reacts with DI/DII of all DENV serotypes.

(A) Neutralization (○) and enhancement (●) of infection by recombinant unmodified mAbs.

(B) Neutralization (○) and enhancement (●) of infection by engineered LALA variant mAbs. Nonlinear regression analysis was used to calculate the  $EC_{50}$ , indicated in the panel. Data are mean of duplicates. One representative experiment out of three performed.

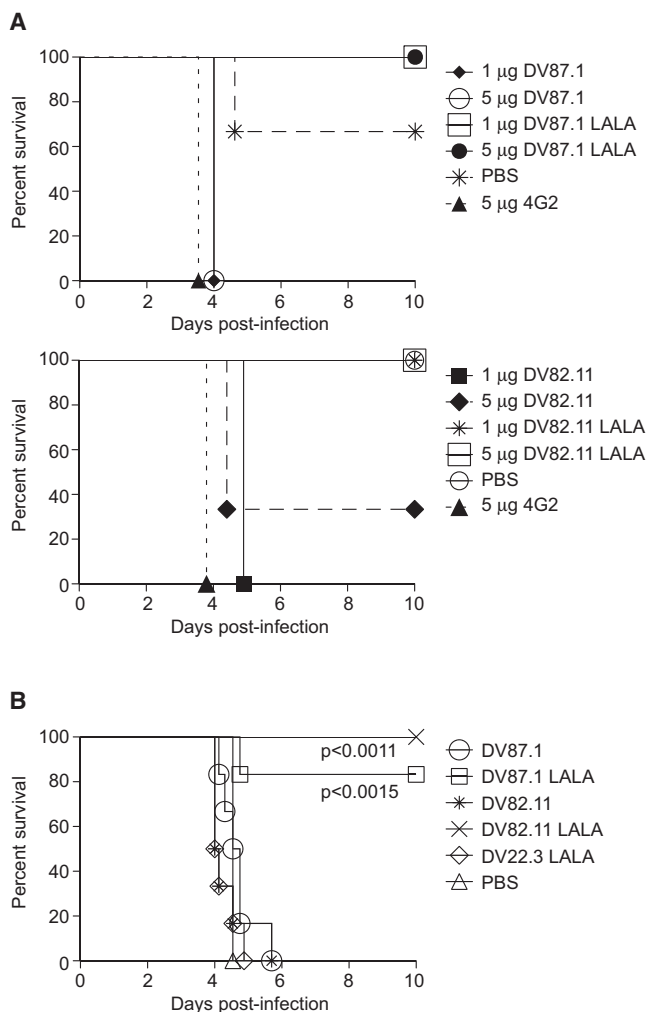
(C) A cocktail of LALA-mAbs (DV82.11 = 2 μg/ml, DV87.1 = DV22.3 = 0.2 μg/ml) was used in combination with mAbs directed against prM (DV69.6) or DI/DII E protein (DV78.6) or serum from a primary DENV-infected donor at potent enhancing concentration (0.1 μg/ml) on K562 cells. Numbers in quadrants indicate the percentage of infected cells, as revealed by staining with an anti-E mouse mAb. The data shown are from one experiment representative of two performed. See also Figure S1 and Table S2.

infection weakly, if at all, while potentially mediating ADE (Falconar, 1999; Henchal et al., 1985; Huang et al., 2006; Men et al., 2004; Roehrig et al., 1998) suggest a potential role for this class of antibodies in enhancing DENV infection in humans. Recent studies in infants have also implicated anti-prM antibodies in the susceptibility of infants to severe dengue (Chau et al., 2009). It has also been proposed that anti-prM antibodies could enhance DENV propagation by enhancing the infectivity of immature, “noninfectious” virions (Rodenhuis-Zybert et al., 2010). However, because the vast majority (>99%) of humans experiencing secondary DENV infection do not develop severe life-threatening

All neutralizing mAbs isolated in this study mediated ADE at subneutralizing concentrations, whereas some E-specific mAbs and most prM-specific mAbs mediated ADE but lacked neutralizing activity. A previous study reported the isolation of three human E protein-specific mAbs, two of which showed enhancing but not neutralizing activity (Schieffelin et al., 2010). More recently, Screaton and coworkers showed that prM-specific mAbs represent a major component of the human DENV-specific response and that these mAbs are highly cross-reactive and, even at high concentrations, do not neutralize infection but potentially promote ADE (Dejnirattisai et al., 2010). Consistent with these data, most of the prM-specific mAbs described in the present study were highly cross-reactive and showed ADE activity over a broad range of concentrations. The human studies together with previous studies on mouse anti-prM mAbs that neutralize

enriching disease, the DI/DII-specific antibodies or the less frequent highly neutralizing DIII-specific antibodies are likely protective, along with other components of the innate and adaptive immune system.

The development of a vaccine against DENV has been problematic, in part due to the possible risk of eliciting suboptimal immune responses that might lead to ADE and severe disease after infection with heterologous virulent strains. In the absence of an effective DENV vaccine, passive immunotherapy with neutralizing antibodies may provide an alternative for the treatment of dengue. A postexposure prophylaxis approach is currently being pursued for treatment of several viral diseases such as respiratory syncytial virus (RSV), rabies, hepatitis B, and WNV (Marasco and Sui, 2007). Humanized anti-DENV mAbs obtained from mice or primates engineered with mutations



**Figure 3. LALA Variants Do Not Enhance DENV In Vivo and Demonstrate Postexposure Therapeutic Efficacy**

(A) (Top) 1 or 5  $\mu$ g of DV87.1, DV87.1 LALA variant, 4G2, or PBS were transferred i.p. in 200  $\mu$ l volume into AG129 mice ( $n = 3$  per group). The mice were infected 18–24 hr later with  $10^6$  pfu DENV-2 strain D2S10. (Bottom) 1 or 5  $\mu$ g of DV82.11, DV82.11 LALA variant, 4G2, or PBS were transferred i.p. in 200  $\mu$ l volume into AG129 mice ( $n = 3$  per group). The mice were subsequently infected 18–24 hr later with  $10^6$  pfu DENV-2 D2S10. All mice receiving LALA-variant mAbs survived as compared to mice receiving a lethal, antibody-enhanced infection (1 or 5  $\mu$ g DV87.1 LALA,  $p < 0.0253$ ; 1 or 5  $\mu$ g DV82.11 LALA,  $p < 0.0455$  compared to 4G2 mAb positive control).

(B) AG129 mice were administered 35  $\mu$ l anti-DENV-1 serum i.p. and were infected 24 hr later with  $10^6$  pfu of DENV-2 D2S10 i.v. At 24 hr after infection, the mice were treated with 50  $\mu$ g of either DV87.1,  $n = 6$ ; DV87.1 LALA,  $n = 6$ ; DV82.11,  $n = 6$ ; DV82.11 LALA,  $n = 5$ ; DV22.3 LALA,  $n = 6$ ; or PBS,  $n = 6$  i.p. In all cases, mortality was monitored daily for 10 days. Mice treated with either 50  $\mu$ g DV87.1 LALA ( $p < 0.0015$ ) or DV82.11 LALA ( $p < 0.011$ ) 24 hr postinfection survived significantly longer than mice receiving 50  $\mu$ g of nonbinding LALA variant mAb DV22.3.

See also Figure S2.

that prevent binding to FcR have been developed for this purpose (Balsitis et al., 2010; Goncalves et al., 2007), but fully human mAbs generated during a natural infection represent an attractive alternative, with a reduced antigenicity and risk of

autoreactivity. Here, we selected three potent human neutralizing mAbs as component of a cocktail capable of targeting two distinct epitopes on each of the four DENV serotypes and engineered them with two leucine-to-alanine mutations in the CH2 region that prevent binding to Fc $\gamma$ R while preserving binding to the neonatal FcRn (Hessell et al., 2007). The LALA variants retained neutralizing activity and, when tested in vitro, failed to enhance DENV infection and inhibited enhancement elicited by polyclonal immune sera or appropriate concentrations of intact antibodies. When tested in vivo in a mouse model of lethal antibody-enhanced vascular leak syndrome, the LALA mAbs protected against lethal infection induced by DENV-immune serum as a postexposure therapeutic. Even though effector functions can enhance protective efficacy of anti-flavivirus antibodies (Chung et al., 2006; Mehlhop et al., 2009; Oliphant et al., 2005; Schlesinger et al., 1993), they are not required for highly neutralizing antibodies. Thus, at least in mice, even after ADE is initiated, there is a time window in which neutralizing antibodies that lack the ability to enhance infection can still protect against lethal disease. These results are a foundation for further investigation to address whether antibody-based therapeutics can control severe DENV disease in humans.

## EXPERIMENTAL PROCEDURES

### Memory B Cell Immortalization and Cloning

Peripheral blood samples were obtained from donors who had been diagnosed with DENV infection. The study protocol was approved by the Scientific and Ethical Committee of the Hospital for Tropical Diseases and the Oxford Tropical Research Ethical Committee. All provided written informed consent. The determination of primary or secondary dengue was made on the basis of acute serology and results of PRNT<sub>50</sub> assays in late convalescence. Peripheral blood mononuclear cells were isolated and cryopreserved. On the day of use, PBMC were thawed and IgG<sup>+</sup> memory B cells were enriched using CD22 microbeads (Miltenyi) followed by cell sorting as described (Traggiai et al., 2004). Cells were immortalized at 20 cells/well in multiple cultures using EBV with CpG oligodeoxynucleotide 2006 (Microsynth) and irradiated allogeneic PBMC. After 2 weeks, the culture supernatants were screened for the presence of DENV-specific mAbs, and positive cultures were cloned by limiting dilution.

### Intracellular Staining

C6/36 or Vero cells were infected with the DENV-3 recovered from donor 76, with the DENV-1 recovered from donor 12, with DENV-2 WHO reference strain 516803, or with DENV-1–4 vaccine strains at MOI leading to 70%–90% of infected cells. After 5 days, cells were harvested, fixed with 2% formaldehyde (Sigma), and permeabilized in PBS 1% FCS 0.5% saponin (Sigma). Cells were then incubated for 1 hr on ice with B cell culture supernatants, washed and stained with Cy5-conjugated goat anti-human IgG (Jackson ImmunoResearch), and analyzed on a FACSArray (BD Biosciences).

### ELISA

96-well ELISA plates were coated at 4°C with recombinant E protein from DENV-1–4 (Rey, 2003), with recombinant DIII from the DENV-3 of donor 76, or with DIII of DENV-4 (Simonelli et al., 2010) (all at 1  $\mu$ g/ml in 0.1M Na carbonate buffer [pH 9.3]). After washing and blocking, mAbs were added for 1 hr at 37°C. After further wash, mAbs bound were revealed using goat anti-human IgG coupled to alkaline phosphatase (Jackson ImmunoResearch). In some experiments, ELISA plates were coated with the 4G2 (1  $\mu$ g/ml) followed by a mixture of DENV-1–4 antigens.

### Yeast Expression and Staining

Yeast expressing DI/DII and DIII of DENV-2 E protein were generated as previously described (see [Supplementary Experimental Procedures](#)) (Oliphant et al., 2006; Sukupolvi-Petty et al., 2007). Yeast cells expressing wild-type or mutant DI/DII or DIII of DENV or WNV (strain New York 1999) E protein were stained with mAbs on ice for 30 min. The cells were then incubated with a goat anti-human IgG conjugated to Cy5 (Jackson ImmunoResearch) for 30 min on ice and analyzed by flow cytometry.

### Immunoblot Analysis

Equal amounts of concentrated supernatant from DENV-3- or DENV-2-infected or uninfected Vero cells were separated under nonreducing conditions by SDS-PAGE and transferred onto nitrocellulose membranes. Filters were blocked with 10% dry skim milk and probed with mAbs. Blots were incubated with horseradish peroxidase-conjugated mouse anti-human IgG, sheep anti-mouse IgG, or donkey anti-rabbit IgG and developed with ECL Plus Western Blotting Detection System (GE Healthcare). E protein was identified using DV55.1, prM protein was identified using the mouse anti-prM mAb 2H2 (Falconar, 1999), and NS1 protein was identified using a rabbit anti-NS1 serum (Young et al., 2000). The C protein was identified as a 16 kD band.

### Production of Recombinant IgG and LALA Variants

RNA was isolated with RNeasy kit (QIAGEN) from EBV-immortalized B cell clones DV82.11, DV87.1, and DV22.3 and cDNA synthesized with M-MLV reverse transcriptase (Invitrogen). Variable regions of heavy-chain and light-chain genes were sequenced and cloned by PCR into human Ig $\gamma$ 1, Ig $\kappa$ , and Ig $\lambda$  expression vectors (provided by M. Nussenzweig, Howard Hughes Medical Institute, The Rockefeller University, New York) using gene-specific primers and Pfu Turbo (Stratagene) (Tiller et al., 2008). The vectors contain a murine Ig gene signal peptide sequence and a multiple cloning site upstream of human Ig $\gamma$ 1, Ig $\kappa$ , and Ig $\lambda$  constant regions, with either wild-type or with leucine-to-alanine mutations at positions CH2 1.3 and 1.2 of Ig $\gamma$ 1, according to the IMGT unique numbering for C-DOMAIN (LALA mutant) (Hessell et al., 2007) introduced by site-directed mutagenesis (GenScript). Equal amounts of heavy- and light-chain vectors were mixed with an equal amount of polyethylenimine (Sigma), incubated for 15 min, and added to 80% confluent HEK293T cells (ATCC) in DMEM supplemented with 1% Nutridoma (Roche). Antibodies were recovered from supernatants harvested 3 days after transfection and purified using protein A affinity chromatography followed by Sephadex 200 size exclusion chromatography.

### Infection Neutralization and Enhancement

DENV neutralization was measured using a flow-based assay. The day before the infection, 5000 Vero cells were plated in 96-well flat-bottom plates. Different dilutions of mAb were mixed with DENV (all at MOI of 0.04) for 1 hr at 37°C and then added to Vero cells. After 3 days, the cells were fixed with 2% formaldehyde, permeabilized in PBS 1% FCS 0.5% saponin, and stained with 4G2. The cells were incubated with a goat anti-mouse IgG conjugated to R-phycoerythrin (Southern Biotechnology) and analyzed by flow cytometry. EC<sub>50</sub> values were determined by nonlinear regression analysis. ADE was measured by a flow assay using K562 cells. mAbs and DENV (MOI of 0.04) were mixed for 1 hr at 37°C and added to 5000 K562/well. After 3 days, cells were fixed, permeabilized, and stained with 4G2, and the number of infected cells was determined by flow cytometry as above.

### Effect of LALA Antibodies on ADE

Potent enhancing antibodies (0.1  $\mu$ g/ml) or serum from primary dengue-infected donors were mixed with an equal volume of LALA cocktail (DV82.11 = 2  $\mu$ g/ml; DV87.1 = DV22.3 = 0.2  $\mu$ g/ml). Attenuated DENV-1–4 virus strains (MOI of 0.04) and the antibody were mixed, added to K562 cells, and incubated for 3 days. Cells were stained with 4G2 and analyzed by flow cytometry as describe above.

### Infection of AG129 Mice

129/Sv mice lacking the interferon  $\alpha/\beta$  and  $\gamma$  receptors (AG129) (van den Broek et al., 1995) were bred in the University of California, Berkeley Animal Facility.

All experimental procedures were preapproved by the University of California Berkeley Animal Care and Use Committee (ACUC) and were performed according to the guidelines of the UC Berkeley ACUC. Mice were injected i.p. with mAb, PBS, or anti-DENV sera in a total volume of 200  $\mu$ l and then infected 18–24 hr later with either 10<sup>5</sup> or 10<sup>6</sup> pfu of DENV strain D2S10 diluted in 100  $\mu$ l volume by intravenous (i.v.) injection into the tail vein. In some experiments, 50  $\mu$ g of mAb were transferred to mice i.v. 24 hr postinfection to assess the therapeutic efficacy of the mAb.

### Statistical Analysis

Mortality data were expressed using Kaplan-Meier survival curves, and log rank analyses were used to determine statistical significance using Prism (GraphPad software). Viral load values for different tissues were compared between experimental groups using a two-sample Wilcoxon rank-sum analysis (Stata v10, College Station, Texas).

### SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, two figures, and two tables and can be found with this article online at [doi:10.1016/j.chom.2010.08.007](https://doi.org/10.1016/j.chom.2010.08.007).

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